Water structure changes in oxime-mediated reactivation process of phosphorylated human acetylcholinesterase

Zueva I., Lushchekina S., Masson P. Kazan Federal University, 420008, Kremlevskaya 18, Kazan, Russia

Abstract

c 2018 The Author(s). The role of water in oxime-mediated reactivation of phosphylated cholinesterases (ChEs) has been asked with recurrence. To investigate oximate water structure changes in this reaction, reactivation of paraoxon-inhibited human acetylcholinesterase (AChE) was performed by the oxime asoxime (HI-6) at different pH in the presence and absence of lyotropic salts: a neutral salt (NaCl), a strong chaotropic salt (LiSCN) and strong kosmotropic salts (ammonium sulphate and phosphate HPO42–). At the same time, molecular dynamic (MD) simulations of enzyme reactivation under the same conditions were performed over 100 ns. Reactivation kinetics showed that the low concentration of chaotropic salt up to 75 mM increased the percentage of reactivation of diethylphosphorylated AChE whereas kosmotropic salts lead only to a small decrease in reactivation. This indicates that water-breaker salt induces de-structuration of water molecules that are electrostricted around oximate ions. Desolvation of oximate favors nucleophilic attack on the phosphorus atom. Effects observed at high salt concentrations (>100 mM) result either from salting-out of the enzyme by kosmotropic salts (phosphate and ammonium sulphate) or denaturing action of chaotropic LiSCN. MDs simulations of diethylphosphorylated hAChE complex with HI-6 over 100 ns were performed in the presence of 100 mM (NH4)2SO4 and 50 mM LiSCN. In the presence of LiSCN, it was found that protein and water have a higher mobility, i.e. water is less organized, compared with the ammonium sulphate system. LiSCN favors protein solvation (hydrophobic hydration) and breakage of elelectrostricted water molecules around of oximate ion. As a result, more free water molecules participated to reaction steps accompanying oxime-mediated dephosphorylation.

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References

- Main, A.R. (1979) Mode of action of anticholinesterases. Pharmacol. Ther. 6, 579-628, https://doi.org/10.1016/0163-7258(79)90066-4
- [2] Masson, P. (2016) Novel approaches in prophylaxis/pretreatment and treatment of organophosphorus poisoning. Phosphorus Sulfur Silicon Relat. Elem. 191, 1433-1443, https://doi.org/10.1080/10426507.2016.1211652
- [3] Mercey, G., Verdelet, T., Renou, J., Kliachyna, M., Baati, R., Nachon, F. et al. (2012) Reactivators of acetylcholinesterase inhibited by organophosphorus nerve agents. Acc. Chem. Res. 45, 756-766, https://doi.org/10.1021/ar2002864
- [4] Mann, T.M., Price, M.E., Whitmore, C.L., Perrott, R.L., Laws, T.R., McColm, R.R. et al. (2017) Bioscavenger is effective as a delayed therapeutic intervention following percutaneous VX poisoning in the guinea-pig. Toxicol. Lett., https://doi.org/10.1016/j.toxlet.2017.11.029

- [5] Sanson, B., Nachon, F., Colletier, J.P., Froment, M.T., Toker, L., Greenblatt, H.M. et al. (2009) Crystallographic snapshots of nonaged and aged conjugates of soman with acetylcholinesterase, and of a ternary complex of the aged conjugate with pralidoxime. J. Med. Chem. 52, 7593–7603, https://doi.org/10.1021/jm900433t
- [6] Carletti, E., Li, H., Li, B., Ekstrom, F., Nicolet, Y., Loiodice, M. et al. (2008) Aging of cholinesterases phosphylated by tabun proceeds through O-dealkylation. J. Am. Chem. Soc. 130, 16011–16020, https://doi.org/10.1021/ja804941z
- [7] Masson, P., Nachon, F. and Lockridge, O. (2010) Structural approach to the aging of phosphylated cholinesterases. Chem. Biol. Interact. 187, 157–162, https://doi.org/10.1016/j.cbi.2010.03.027
- [8] Quinn, D.M., Topczewski, J., Yasapala, N. and Lodge, A. (2017) Why is aged acetylcholinesterase so difficult to reactivate? Molecules 22, 1464, https://doi.org/10.3390/molecules22091464
- [9] Luo, C., Saxena, A., Smith, M., Garcia, G., Radic, Z., Taylor, P. et al. (1999) Phosphoryl oxime inhibition of acetylcholinesterase during oxime reactivation is prevented by edrophonium. Biochemistry 38, 9937–9947, https://doi.org/10.1021/bi9905720
- [10] Davies, D.R. and Green, A.L. (1956) The kinetics of reactivation, by oximes, of cholinesterase inhibited by organophosphorus compounds. Biochem. J. 63, 529–535, https://doi.org/10.1042/bj0630529
- [11] Wang, E.I. and Braid, P.E. (1967) Oxime reactivation of diethylphosphoryl human serum cholinesterase. J. Biol. Chem. 242, 2683–2687
- [12] Driant, T., Nachon, F., Ollivier, C., Renard, P.Y. and Derat, E. (2017) On the influence of the protonation states of active site residues on AChE reactivation: a QM/MM approach. ChemBiochem 18, 666-675, https://doi.org/10.1002/cbic.201600646
- [13] Berman, H.A. (2005) Rate enhancements and the mechanism of oxime-mediated reactivation. Chem. Biol. Interact. 157–158, 356–357, https://doi.org/10.1016/j.cbi.2005.10.048
- [14] Koellner, G., Kryger, G., Millard, C.B., Silman, I., Sussman, J.L. and Steiner, T. (2000) Active-site gorge and buried water molecules in crystal structures of acetylcholinesterase from Torpedo californica. J. Mol. Biol. 296, 713-735, https://doi.org/10.1006/jmbi.1999.3468
- [15] Masson, P., Clery, C., Guerra, P., Redslob, A., Albaret, C. and Fortier, P.L. (1999) Hydration change during the aging of phosphorylated human butyrylcholinesterase: importance of residues aspartate-70 and glutamate-197 in the water network as probed by hydrostatic and osmotic pressures. Biochem. J. 343, 361–369, https://doi.org/10.1042/bj3430361
- [16] Nachon, F., Asojo, O.A., Borgstahl, G., Masson, P. and Lockridge, O. (2005) Structural data on the aging of diethylphosphoryl-butyrylcholinesterase. Chem. Biol. Interact. 157–158, 408–409, https://doi.org/10.1016/j.cbi.2005.10.078
- [17] Masson, P., Lushchekina, S., Schopfer, L.M. and Lockridge, O. (2013) Effects of viscosity and osmotic stress on the reaction of human butyrylcholinesterase with cresyl saligenin phosphate, a toxicant related to the aerotoxic syndrome: kinetic and molecular dynamics studies. Biochem. J. 454, 387-399, https://doi.org/10.1042/BJ20130389
- [18] Peters, J., Martinez, N., Trovaslet, M., Scannapieco, K., Koza, M.M., Masson, P. et al. (2016) Dynamics of human acetylcholinesterase bound to non-covalent and covalent inhibitors shedding light on changes to the water network structure. Phys. Chem. Chem. Phys. 18, 12992–13001, https://doi.org/10.1039/C6CP00280C
- [19] Zhang, Y. and Cremer, P.S. (2006) Interactions between macromolecules and ions: the hofmeister series. Curr. Opin. Chem. Biol. 10, 658-663, https://doi.org/10.1016/j.cbpa.2006.09.020
- [20] Leuzinger, W. (1971) The number of catalytic sites in acetylcholinesterase. Biochem. J. 123, 139–141, https://doi.org/10.1042/bj1230139
- [21] Ekström, F., Hörnberg, A., Artursson, E., Hammarström, L.-G., Schneider, G. and Pang, Y.-P. (2009) Structure of HI-6•Sarin-Acetylcholinesterase determined by X-Ray crystallography and molecular dynamics simulation: reactivator mechanism and design. PLoS ONE 4, e5957, https://doi.org/10.1371/journal.pone.0005957
- [22] Ashani, Y., Radic, Z., Tsigelny, I., Vellom, D.C., Pickering, N.A., Quinn, D.M. et al. (1995) Amino acid residues controlling reactivation of organophosphonyl conjugates of acetylcholinesterase by mono- and bisquaternary oximes. J. Biol. Chem. 270, 6370–6380, https://doi.org/10.1074/jbc.270.11.6370
- [23] Terrier, F., Rodriguez-Dafonte, P., Le Guevel, E. and Moutiers, G. (2006) Revisiting the reactivity of oximate alpha-nucleophiles with electrophilic phosphorus centers. Relevance to detoxification of sarin, soman and DFP under mild conditions. Org. Biomol. Chem. 4, 4352–4363, https://doi.org/10.1039/B609658C
- [24] Worek, F., Thiermann, H., Szinicz, L. and Eyer, P. (2004) Kinetic analysis of interactions between human acetylcholinesterase, structurally different organophosphorus compounds and oximes. Biochem. Pharmacol. 68, 2237–2248, https://doi.org/10.1016/j.bcp.2004.07.038
- [25] Macek Hrvat, N., Zorbaz, T., Sinko, G. and Kovarik, Z. (2017) The estimation of oxime efficiency is affected by the experimental design of phosphylated acetylcholinesterase reactivation. Toxicol. Lett. 293, 222–228
- [26] Ellman, G.L., Courtney, K.D., Andres, V. and Feather-Stone, R.M. (1961) A new and rapid colorimetric determination of acetylcholinesterase activity. Biochem. Pharmacol. 7, 88–95, https://doi.org/10.1016/0006-2952(61)90145-9

- [27] Cheung, J., Rudolph, M.J., Burshteyn, F., Cassidy, M.S., Gary, E.N., Love, J. et al. (2012) Structures of human acetylcholinesterase in complex with pharmacologically important ligands. J. Med. Chem. 55, 10282–10286, https://doi.org/10.1021/jm300871x
- [28] Pedretti, A., Villa, L. and Vistoli, G. (2004) VEGA An open platform to develop chemo-bio-informatics applications, using plug-in architecture and script programming. J. Comput. Aided Mol. Des. 18, 167–173, https://doi.org/10.1023/B:JCAM.0000035186.90683.f2
- [29] Vanommeslaeghe, K., Hatcher, E., Acharya, C., Kundu, S., Zhong, S., Shim, J. et al. (2010) CHARMM general force field: a force field for drug-like molecules compatible with the CHARMM all-atom additive biological force fields. J. Comput. Chem. 31, 671-690
- [30] Vanommeslaeghe, K. and MacKerell, Jr, A.D. (2012) Automation of the CHARMM General Force Field (CGenFF) I: bond perception and atom typing. J. Chem. Inf. Model. 52, 3144–3154, https://doi.org/10.1021/ci300363c
- [31] Vanommeslaeghe, K., Raman, E.P. and MacKerell, Jr, A.D. (2012) Automation of the CHARMM General Force Field (CGenFF) II: assignment of bonded parameters and partial atomic charges. J. Chem. Inf. Model. 52, 3155-3168, https://doi.org/10.1021/ci3003649
- [32] Zhu, X., Lopes, P. E.M. and MacKerell, A.D. (2011) Recent developments and applications of the CHARMM force fields. Wiley Interdisciplinary Reviews: Computational Molecular Science, John Wiley & Sons, Inc
- [33] Mayne, C.G., Saam, J., Schulten, K., Tajkhorshid, E. and Gumbart, J.C. (2013) Rapid parameterization of small molecules using the force field toolkit. J. Comput. Chem. 34, 2757–2770, https://doi.org/10.1002/jcc.23422
- [34] Levine, B.G., Stone, J.E. and Kohlmeyer, A. (2011) Fast analysis of molecular dynamics trajectories with graphics processing units-radial distribution function histogramming. J. Comput. Phys. 230, 3556–3569, https://doi.org/10.1016/j.jcp.2011.01.048
- [35] Phillips, J.C., Braun, R., Wang, W., Gumbart, J., Tajkhorshid, E., Villa, E. et al. (2005) Scalable molecular dynamics with NAMD. J. Comput. Chem. 26, 1781–1802, https://doi.org/10.1002/jcc.20289
- [36] Best, R.B., Zhu, X., Shim, J., Lopes, P.E.M., Mittal, J., Feig, M. et al. (2012) Optimization of the additive CHARMM all-atom protein force field targeting improved sampling of the backbone φ, ψ and side-chain χ1 and χ2 dihedral angles. J. Chem. Theory Comput. 8, 3257–3273, https://doi.org/10.1021/ct300400x
- [37] Sadovnichy, V., Tikhonravov, A., Voevodin, V. and Opanasenko, V. (2013) "Lomonosov": supercomputing at Moscow State University. In Contemporary High Performance Computing: From Petascale toward Exascale (Vetter, J.S., ed.), pp. 283–307, CRC Press, Boca Raton, U.S.A.
- [38] Humphrey, W., Dalke, A. and Schulten, K. (1996) VMD: visual molecular dynamics. J. Mol. Graph. 14, 33–38, https://doi.org/10.1016/0263-7855(96)00018-5
- [39] Bakan, A., Meireles, L.M. and Bahar, I. (2011) ProDy: protein dynamics inferred from theory and experiments. Bioinformatics 27, 1575–1577, https://doi.org/10.1093/bioinformatics/btr168
- [40] Worek, F., Wille, T., Koller, M. and Thiermann, H. (2012) Reactivation kinetics of a series of related bispyridinium oximes with organophosphate-inhibited human acetylcholinesterase—Structure-activity relationships. Biochem. Pharmacol. 83, 1700-1706, https://doi.org/10.1016/j.bcp.2012.03.002
- [41] Cacace, M.G., Landau, E.M. and Ramsden, J.J. (1997) The Hofmeister series: salt and solvent effects on interfacial phenomena. Q. Rev. Biophys. 30, 241–277, https://doi.org/10.1017/S0033583597003363
- [42] Broering, J.M. and Bommarius, A.S. (2005) Evaluation of Hofmeister effects on the kinetic stability of proteins. J. Phys. Chem. B 109, 20612–20619, https://doi.org/10.1021/jp053618+
- [43] Wang, I.C. and Braid, P.E. (1977) Compensational phenomena in reactivation of dimethyland diethylphosphoryl butyrylcholinesterases. Biochim. Biophys. Acta 481, 515–525, https://doi.org/10.1016/0005-2744(77)90284-4
- [44] Moutiers, G., Le Guével, E., Villien, L. and Terrier, F. (1997) Similar catalytic behaviour of oximate and phenoxide bases in the ionization of bis(2,4-dinitrophenyl)methane in 50% water- 50% Me2SO. Revisiting the role of solvational imbalances in determining the nucleophilic reactivity of α-effect oximate bases. J. Chem. Soc. Perkin Trans. 2, 7-14, https://doi.org/10.1039/a605249e
- [45] Buncel, E., Cannes, C., Chatrousse, A.-P. and Terrier, F. (2002) Reactions of oximate α-nucleophiles with esters: evidence from solvation effects for substantial decoupling of desolvation and bond formation. J. Am. Chem. Soc. 124, 8766–8767, https://doi.org/10.1021/ja020379k