

Bacterial ribonuclease: Mutagenic effect in microbial test-systems

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Abstract

Pure enzyme samples of ribonuclease from *Bacillus intermedius* 7P (known commercially as 'binase') were investigated for genotoxicity in four microbial tests: the Ames plate incorporation method, Ara(R)-assay; the prophage induction test; and the DNA-repair test. The weak mutagenic effect of binase at high concentrations (0.1 mg/plate, 1 mg/plate) was established by induction of forward Ara(R)-mutations and histidine-reverse mutations (both frameshift mutations and base pair substitution). Metabolic activation with rat or chicken liver, human placenta or plant (from tulip bulbs) microsomal fractions in vitro was seen to abolish the binase mutagenicity. *Bacillus intermedius* 7P ribonuclease appears to possess DNA damaging activity in *uvrA*- and *polA*- mutants, but not in the *recA*-deficient *Escherichia coli* strain, and exhibits an induction of *recA*-dependent mutagenesis detected by the 8-fold increase of the prophage-induction level in lysogenic *Bacillus subtilis* culture and by the 5-fold increase of this level in the *Streptomyces lavendulae* 3 lysogenic strain. The importance of the roles of both of enzyme catalytic activity and native structure is emphasized. A proposed mechanism for exogenous ribonuclease action is discussed. *Bacillus intermedius* 7P ribonuclease probably does not act as a direct genotoxic agent interacting with DNA, but could provoke nucleotide imbalance through its catalytic action on membrane-associated RNAs, which results in alteration of DNA replication and, as a consequence, in *recA*-dependent mutagenesis.
