

Exposure to the Epstein-Barr viral antigen latent membrane protein 1 induces myelin-reactive antibodies in vivo

Lomakin Y., Arapidi G., Chernov A., Ziganshin R., Tcyganov E., Lyadova I., Butenko I., Osetrova M., Ponomarenko N., Telegin G., Govorun V., Gabibov A., Belogurov A.

Kazan Federal University, 420008, Kremlevskaya 18, Kazan, Russia

Abstract

© 2017 Lomakin, Arapidi, Chernov, Ziganshin, Tcyganov, Lyadova, Butenko, Osetrova, Ponomarenko, Telegin, Govorun, Gabibov and Belogurov. Multiple sclerosis (MS) is an autoimmune chronic inflammatory disease of the central nervous system (CNS). Cross-reactivity of neuronal proteins with exogenous antigens is considered one of the possible mechanisms of MS triggering. Previously, we showed that monoclonal myelin basic protein (MBP)-specific antibodies from MS patients cross-react with Epstein-Barr virus (EBV) latent membrane protein 1 (LMP1). In this study, we report that exposure of mice to LMP1 results in induction of myelin-reactive autoantibodies in vivo. We posit that chronic exposure or multiple acute exposures to viral antigen may redirect B cells from production of antiviral antibodies to antibodies, specific to myelin antigen. However, even in inbred animals, which are almost identical in terms of their genomes, such an effect is only observed in 20-50% of animals, indicating that this change occurs by chance, rather than systematically. Cross-immunoprecipitation analysis showed that only part of anti-MBP antibodies from LMP1-immunized mice might simultaneously bind LMP1. In contrast, the majority of anti-LMP1 antibodies from MBP-immunized mice bind MBP. De novo sequencing of anti-LMP1 and anti-MBP antibodies by mass spectrometry demonstrated enhanced clonal diversity in LMP1-immunized mice in comparison with MBP-immunized mice. We suggest that induction of MBP-reactive antibodies in LMP1-immunized mice may be caused by either Follicular dendritic cells (FDCs) or by T cells that are primed by myelin antigens directly in CNS. Our findings help to elucidate the still enigmatic link between EBV infection and MS development, suggesting that myelin-reactive antibodies raised as a response toward EBV protein LMP1 are not truly cross-reactive but are primarily caused by epitope spreading.

<http://dx.doi.org/10.3389/fimmu.2017.00777>

Keywords

Autoantibodies, Cross-reactivity, Epitope spreading, Epstein-Barr virus, Latent membrane protein 1, Multiple sclerosis, Myelin basic protein

References

- [1] Beasley SJ, Greer JM. Autoantibodies and their potential roles in diseases of the nervous system. *Clin Exp Neuroimmunol* (2015) 6(4):370-86. doi:10.1111/cen3.12269

- [2] Sandel PC, Monroe JG. Negative selection of immature B cells by receptor editing or deletion is determined by site of antigen encounter. *Immunity* (1999) 10(3):289-99. doi:10.1016/S1074-7613(00)80029-1
- [3] Pelanda R, Torres RM. Central B-cell tolerance: where selection begins. *Cold Spring Harb Perspect Biol* (2012) 4(4):a007146. doi:10.1101/cshperspect.a007146
- [4] Coutinho A, Kazatchkine MD, Avrameas S. Natural autoantibodies. *Curr Opin Immunol* (1995) 7(6):812-8. doi:10.1016/0952-7915(95)80053-0
- [5] Quintana FJ, Cohen IR. The natural autoantibody repertoire and autoimmune disease. *Biomed Pharmacother* (2004) 58(5):276-81. doi:10.1016/j.biopha.2004.04.011
- [6] Madi A, Hecht I, Bransburg-Zabary S, Merbl Y, Pick A, Zucker-Toledano M, et al. Organization of the autoantibody repertoire in healthy newborns and adults revealed by system level informatics of antigen microarray data. *Proc Natl Acad Sci U S A* (2009) 106(34):14484-9. doi:10.1073/pnas.0901528106
- [7] Xing Y, Li W, Lin Y, Fu M, Li CX, Zhang P, et al. The influence of BCR density on the differentiation of natural poly-reactive B cells begins at an early stage of B cell development. *Mol Immunol* (2009) 46(6):1120-8. doi:10.1016/j.molimm.2008.10.031
- [8] Daffa NI, Tighe PJ, Corne JM, Fairclough LC, Todd I. Natural and disease-specific autoantibodies in chronic obstructive pulmonary disease. *Clin Exp Immunol* (2015) 180(1):155-63. doi:10.1111/cei.12565
- [9] Lacroix-Desmazes S, Kaveri SV, Mouthon L, Ayoub A, Malanchere E, Coutinho A, et al. Self-reactive antibodies (natural autoantibodies) in healthy individuals. *J Immunol Methods* (1998) 216(1-2):117-37. doi:10.1016/S0022-1759(98)00074-X
- [10] Dimitrov JD, Planchais C, Roumenina LT, Vassilev TL, Kaveri SV, Lacroix-Desmazes S. Antibody polyreactivity in health and disease: statu variabilis. *J Immunol* (2013) 191(3):993-9. doi:10.4049/jimmunol.1300880
- [11] Zhang W, Nardi MA, Borkowsky W, Li Z, Karpatkin S. Role of molecular mimicry of hepatitis C virus protein with platelet GPIIIa in hepatitis C-related immunologic thrombocytopenia. *Blood* (2009) 113(17):4086-93. doi:10.1182/blood-2008-09-181073
- [12] Li Z, Nardi MA, Karpatkin S. Role of molecular mimicry to HIV-1 peptides in HIV-1-related immunologic thrombocytopenia. *Blood* (2005) 106(2):572-6. doi:10.1182/blood-2005-01-0243
- [13] Kampylafka EI, Alexopoulos H, El Hamidieh A, Dalakas MC, Andreakos E, Tzioufas AG. Immunization of mice with a peptide derived from the HTLV-1 TAX1BP1 protein induces cross-reactive antibodies against aquaporin 4. *Autoimmunity* (2015) 48:453-9. doi:10.3109/08916934.2015.1070836
- [14] Ang CW, Yuki N, Jacobs BC, Koga M, Van Doorn PA, Schmitz PI, et al. Rapidly progressive, predominantly motor Guillain-Barre syndrome with anti-GalNAc-GD1a antibodies. *Neurology* (1999) 53(9):2122-7. doi:10.1212/WNL.53.9.2122
- [15] Steelman AJ. Infection as an environmental trigger of multiple sclerosis disease exacerbation. *Front Immunol* (2015) 6:520. doi:10.3389/fimmu.2015.00520
- [16] Kakalacheva K, Munz C, Lunemann JD. Viral triggers of multiple sclerosis. *Biochim Biophys Acta* (2011) 1812(2):132-40. doi:10.1016/j.bbadis.2010.06.012
- [17] Gabibov AG, Belogurov AA Jr, Lomakin YA, Zakharova MY, Avakyan ME, Dubrovskaya VV, et al. Combinatorial antibody library from multiple sclerosis patients reveals antibodies that cross-react with myelin basic protein and EBV antigen. *FASEB J* (2011) 25(12):4211-21. doi:10.1096/fj.11-190769
- [18] Lomakin YA, Zakharova MY, Belogurov AA, Bykova NA, Dronina MA, Tupikin AE, et al. Polyreactive monoclonal autoantibodies in multiple sclerosis: functional selection from phage display library and characterization by deep sequencing analysis. *Acta Naturae* (2013) 5(4):94-104.
- [19] Lomakin YA, Zakharova MY, Stepanov AV, Dronina MA, Smirnov IV, Bobik TV, et al. Heavy-light chain interrelations of MS-associated immunoglobulins probed by deep sequencing and rational variation. *Mol Immunol* (2014) 62(2):305-14. doi:10.1016/j.molimm.2014.01.013
- [20] Belogurov AA Jr, Kurkova IN, Friboulet A, Thomas D, Misikov VK, Zakharova MY, et al. Recognition and degradation of myelin basic protein peptides by serum autoantibodies: novel biomarker for multiple sclerosis. *J Immunol* (2008) 180(2):1258-67. doi:10.4049/jimmunol.180.2.1258
- [21] Miller SD, Karpus WJ. Experimental autoimmune encephalomyelitis in the mouse. *Curr Protoc Immunol* (2007) Chapter 15:Unit15.1.
- [22] UniProt C. UniProt: a hub for protein information. *Nucleic Acids Res* (2015) 43(Database issue):D204-12. doi:10.1093/nar/gku989
- [23] Lefranc MP, Giudicelli V, Ginestoux C, Jabado-Michaloud J, Folch G, Bellahcene F, et al. IMGT, the international ImMunoGeneTics information system. *Nucleic Acids Res* (2009) 37(Database issue):D1006-12. doi:10.1093/nar/gkn838
- [24] Okonechnikov K, Golosova O, Fursov M, UGENE team. Unipro UGENE: a unified bioinformatics toolkit. *Bioinformatics* (2012) 28(8):1166-7. doi:10.1093/bioinformatics/bts091
- [25] Crooks GE, Hon G, Chandonia JM, Brenner SE. WebLogo: a sequence logo generator. *Genome Res* (2004) 14(6):1188-90. doi:10.1101/gr.849004

- [26] Handel AE, Williamson AJ, Disanto G, Handunnetthi L, Giovannoni G, Ramagopalan SV. An updated meta-analysis of risk of multiple sclerosis following infectious mononucleosis. *PLoS One* (2010) 5(9):e12496. doi:10.1371/journal.pone.0012496
- [27] Lossius A, Riise T, Pugliatti M, Bjornevik K, Casetta I, Drulovic J, et al. Season of infectious mononucleosis and risk of multiple sclerosis at different latitudes; the EnvIMS Study. *Mult Scler* (2014) 20(6):669-74. doi:10.1177/1352458513505693
- [28] El Shikh ME, El Sayed RM, Szakal AK, Tew JG. T-independent antibody responses to T-dependent antigens: a novel follicular dendritic cell-dependent activity. *J Immunol* (2009) 182(6):3482-91. doi:10.4049/jimmunol.0802317
- [29] McMahon EJ, Bailey SL, Castenada CV, Waldner H, Miller SD. Epitope spreading initiates in the CNS in two mouse models of multiple sclerosis. *Nat Med* (2005) 11(3):335-9. doi:10.1038/nm1202
- [30] Zouali M. *Natural Antibodies*. eLS. New Jersey: John Wiley & Sons, Ltd (2015).
- [31] Panda S, Ding JL. Natural antibodies bridge innate and adaptive immunity. *J Immunol* (2015) 194(1):13-20. doi:10.4049/jimmunol.1400844
- [32] Rapaka RR, Ricks DM, Alcorn JF, Chen K, Khader SA, Zheng M, et al. Conserved natural IgM antibodies mediate innate and adaptive immunity against the opportunistic fungus *Pneumocystis murina*. *J Exp Med* (2010) 207(13):2907-19. doi:10.1084/jem.20100034
- [33] Xu JL, Davis MM. Diversity in the CDR3 region of V(H) is sufficient for most antibody specificities. *Immunity* (2000) 13(1):37-45. doi:10.1016/S1074-7613(00)00006-6
- [34] Ippolito GC, Schelonka RL, Zemlin M, Ivanov II, Kobayashi R, Zemlin C, et al. Forced usage of positively charged amino acids in immunoglobulin CDR-H3 impairs B cell development and antibody production. *J Exp Med* (2006) 203(6):1567-78. doi:10.1084/jem.20052217
- [35] Georgiou G, Ippolito GC, Beausang J, Busse CE, Wardemann H, Quake SR. The promise and challenge of high-throughput sequencing of the antibody repertoire. *Nat Biotechnol* (2014) 32(2):158-68. doi:10.1038/nbt.2782