

# ZIKV infection regulates inflammasomes pathway for replication in monocytes

Khaiboullina S., Uppal T., Sarkar R., Goryalski A., St Jeor S., Verma S.

Kazan Federal University, 420008, Kremlevskaya 18, Kazan, Russia

---

## Abstract

© 2017 The Author(s). ZIKV causes microcephaly by crossing the placental barrier, however, the mechanism of trans-placental dissemination of ZIKV remains unknown. Here, we sought to determine whether monocytes, which can cross tissue barriers, assist ZIKV dissemination to the fetus. We determined this by infecting monocytes with two strains of ZIKV: South American (PRVABC59) and Nigerian (IBH30656) and analyzing viral replication. We found that ZIKV infects and replicates in monocytes and macrophages, which results in the modulation of a large number of cellular genes. Analysis of these genes identified multiple pathways including inflammasome to be targeted by ZIKV, which was confirmed by analyzing the transcript levels of the proteins of inflammasome pathways, NLRP3, ASC, caspase 1, IL-1 and IL-18. Interestingly, IFN $\alpha$  and the IFN inducible gene, MxA were not enhanced, suggesting prevention of innate antiviral defense by ZIKV. Also, inhibition of inflammasome led to an increased transcriptional activity of IFN $\alpha$ , MxA and CXCL10. Based on these results we suggest that ZIKV transcription is regulated by inflammasomes.

<http://dx.doi.org/10.1038/s41598-017-16072-3>

---

## References

- [1] MOH. Increase of microcephaly in the northeast of Brazil, <http://moh.gov.jm/epidemiological-alert-increase-of-microcephaly-in-the-northeast-of-brazil/> (2015).
- [2] Campos, G. S., Bandeira, A. C. & Sardi, S. I. Zika Virus Outbreak, Bahia, Brazil. *Emerg Infect Dis* 21, 1885-1886, <https://doi.org/10.3201/eid2110.150847> (2015).
- [3] Dowall, S. D. et al. A Susceptible Mouse Model for Zika Virus Infection. *PLoS Negl Trop Dis* 10, e0004658, <https://doi.org/10.1371/journal.pntd.0004658.PNTD-D-16-00358> (2016).
- [4] Wu, K. Y. et al. Vertical transmission of Zika virus targeting the radial glial cells affects cortex development of offspring mice. *Cell Res* 26, 645-654, <https://doi.org/10.1038/cr.2016.58> (2016).
- [5] Li, C. et al. Zika Virus Disrupts Neural Progenitor Development and Leads to Microcephaly in Mice. *Cell Stem Cell* 19, 120-126, <https://doi.org/10.1016/j.stem.2016.04.017> (2016).
- [6] Cugola, F. R. et al. The Brazilian Zika virus strain causes birth defects in experimental models. *Nature* (2016).
- [7] Maestre, A. M. & Fernandez-Sesma, A. Finding Clues for Congenital Zika Syndrome: Zika Virus Selective Infection of Immature Neurons. *EBioMedicine* 10, 7-8, <https://doi.org/10.1016/j.ebiom.2016.07.026> (2016).
- [8] Tang, H. et al. Zika virus infects human cortical neural progenitors and attenuates their growth. *Cell Stem Cell* 18, 587-590 (2016).
- [9] Garcez, P. P. et al. Zika virus impairs growth in human neurospheres and brain organoids. *Science* 352, 816-818 (2016).
- [10] Bayer, A. et al. Type III interferons produced by human placental trophoblasts confer protection against Zika virus infection. *Cell host & microbe* 19, 705-712 (2016).

- [11] Lazear, H. M. et al. A mouse model of Zika virus pathogenesis. *Cell host & microbe* 19, 720-730 (2016).
- [12] Rossi, S. L. et al. Characterization of a novel murine model to study Zika virus. *The American journal of tropical medicine and hygiene* 94, 1362-1369 (2016).
- [13] Cao, W. et al. Rapid differentiation of monocytes into type I IFN-producing myeloid dendritic cells as an antiviral strategy against influenza virus infection. *The Journal of Immunology* 189, 2257-2265 (2012).
- [14] Hamel, R. et al. Biology of Zika Virus Infection in Human Skin Cells. *Journal of Virology* 89, 8880-8896, <https://doi.org/10.1128/JVI.00354-15> (2015).
- [15] Authors' correction for Euro Surveill. 2016;21(26). *Euro Surveill* 21, <https://doi.org/10.2807/1560-7917.ES.2016.21.31.30305> (2016).
- [16] Gourinat, A. C., O'Connor, O., Calvez, E., Goarant, C. & Dupont-Rouzeyrol, M. Detection of Zika virus in urine. *Emerg Infect Dis* 21, 84-86, <https://doi.org/10.3201/eid2101.140894> (2015).
- [17] Nicastri, E. et al. Persistent detection of Zika virus RNA in semen for six months after symptom onset in a traveller returning from Haiti to Italy, February 2016. *Euro Surveill* 21, <https://doi.org/10.2807/1560-7917.ES.2016.21.32.30314> (2016).
- [18] Bingham, A. M. et al. Comparison of Test Results for Zika Virus RNA in Urine, Serum, and Saliva Specimens from Persons with Travel-Associated Zika Virus Disease-Florida, 2016. *MMWR Morb Mortal Wkly Rep* 65, 475-478, <https://doi.org/10.15585/mmwr.mm6518e2> (2016).
- [19] Arias, R. A., Munoz, L. D. & Munoz-Fernandez, M. A. Transmission of HIV-1 infection between trophoblast placental cells and T-cells take place via an LFA-1-mediated cell to cell contact. *Virology* 307, 266-277, S0042682202000405 [pii] (2003).
- [20] Pereira, L., Maidji, E., McDonagh, S., Genbacev, O. & Fisher, S. Human cytomegalovirus transmission from the uterus to the placenta correlates with the presence of pathogenic bacteria and maternal immunity. *J Virol* 77, 13301-13314 (2003).
- [21] McDonagh, S. et al. Viral and bacterial pathogens at the maternal-fetal interface. *J Infect Dis* 190, 826-834, <https://doi.org/10.1086/422330> JID32235 (2004).
- [22] Parekh, F. K., Davison, B. B., Gamboa, D., Hernandez, J. & Branch, O. H. Placental histopathologic changes associated with subclinical malaria infection and its impact on the fetal environment. *Am J Trop Med Hyg* 83, 973-980, <https://doi.org/10.4269/ajtmh.2010.09-0445> (2010).
- [23] Karniychuk, U. U. & Nauwynck, H. J. Pathogenesis and prevention of placental and transplacental porcine reproductive and respiratory syndrome virus infection. *Vet Res* 44, 95, <https://doi.org/10.1186/1297-9716-4-95> (2013).
- [24] Al-Husaini, A. M. Role of placenta in the vertical transmission of human immunodeficiency virus. *J Perinatol* 29, 331-336, <https://doi.org/10.1038/jp.2008.187> (2009).
- [25] Maciejewski, J. P. et al. Infection of hematopoietic progenitor cells by human cytomegalovirus. *Blood* 80, 170-178 (1992).
- [26] Maciejewski, J. P. et al. Infection of mononucleated phagocytes with human cytomegalovirus. *Virology* 195, 327-336 (1993).
- [27] Pirhonen, J., Sareneva, T., Kurimoto, M., Julkunen, I. & Matikainen, S. Virus infection activates IL-1 $\beta$  and IL-18 production in human macrophages by a caspase-1-dependent pathway. *The Journal of Immunology* 162, 7322-7329 (1999).
- [28] Netea, M. G., van de Veerdonk, F. L., van der Meer, J. W. M., Dinarello, C. A. & Joosten, L. A. B. Inflammasome-Independent Regulation of IL-1-Family Cytokines. *Annual Review of Immunology* 33, 49-77, <https://doi.org/10.1146/annurevimmunol-032414-112306> (2015).
- [29] Brough, D. & Rothwell, N. J. Caspase-1-dependent processing of pro-interleukin-1 $\beta$  is cytosolic and precedes cell death. *J Cell Sci* 120, 772-781, <https://doi.org/10.1242/jcs.03377> (2007).
- [30] Netea, M. G., van de Veerdonk, F. L., van der Meer, J. W., Dinarello, C. A. & Joosten, L. A. Inflammasome-independent regulation of IL-1-family cytokines. *Annu Rev Immunol* 33, 49-77, <https://doi.org/10.1146/annurev-immunol-032414-112306> (2015).
- [31] Henry, C. M. et al. Neutrophil-Derived Proteases Escalate Inflammation through Activation of IL-36 Family Cytokines. *Cell Rep* 14, 708-722, <https://doi.org/10.1016/j.celrep.2015.12.072> (2016).
- [32] Sims, J. E. & Smith, D. E. The IL-1 family: regulators of immunity. *Nat Rev Immunol* 10, 89-102, <https://doi.org/10.1038/nri2691> (2010).
- [33] Afonina, I. S., Muller, C., Martin, S. J. & Beyaert, R. Proteolytic Processing of Interleukin-1 Family Cytokines: Variations on a Common Theme. *Immunity* 42, 991-1004, <https://doi.org/10.1016/j.immuni.2015.06.003> (2015).
- [34] Dinarello, C. A. Immunological and inflammatory functions of the interleukin-1 family. *Annu Rev Immunol* 27, 519-550, <https://doi.org/10.1146/annurev.immunol.021908.132612> (2009).
- [35] Sakaguchi, S., Yamaguchi, T., Nomura, T. & Ono, M. Regulatory T cells and immune tolerance. *Cell* 133, 775-787, <https://doi.org/10.1016/j.cell.2008.05.009> (2008).

- [36] Ferreri, A. J., Illerhaus, G., Zucca, E. & Cavalli, F. Flows and flaws in primary central nervous system lymphoma. *Nat Rev Clin Oncol* 7, 10 1038/nrclinonc 20101039-c1031; author reply 1010:1038/nrclinonc 2010 1039-c1032, 10. 1038/nrclinonc. 2010. 9-c1 (2010).
- [37] Carson, W. E. et al. Coadministration of interleukin-18 and interleukin-12 induces a fatal inflammatory response in mice: critical role of natural killer cell interferon-gamma production and STAT-mediated signal transduction. *Blood* 96, 1465-1473 (2000).
- [38] Ziegler-Heitbrock, H. W. & Ulevitch, R. J. CD14: cell surface receptor and differentiation marker. *Immunol Today* 14, 121-125, [https://doi.org/10.1016/0167-5699\(93\)90212-4](https://doi.org/10.1016/0167-5699(93)90212-4) (1993).
- [39] Uchide, N., Ohyama, K., Yuan, B., Bessho, T. & Yamakawa, T. Differentiation of monocytes to macrophages induced by influenza virus-infected apoptotic cells. *J Gen Virol* 83, 747-751, <https://doi.org/10.1099/0022-1317-83-4-747> (2002).
- [40] Yang, J., Zhang, L., Yu, C., Yang, X. F. & Wang, H. Monocyte and macrophage differentiation: circulation inflammatory monocyte as biomarker for inflammatory diseases. *Biomark Res* 2, 1, <https://doi.org/10.1186/2050-7771-2-1> (2014).
- [41] Downham, E. et al. A novel mitochondrial ND5 (MTND5) gene mutation giving isolated exercise intolerance. *Neuromuscul Disord* 18, 310-314, <https://doi.org/10.1016/j.nmd.2008.01.003> (2008).
- [42] Ahari, S. E. et al. Investigation on mitochondrial tRNA(Leu/Lys), NDI and ATPase 6/8 in iranian multiple sclerosis patients. *Cellular and Molecular Neurobiology* 27, 695-700, <https://doi.org/10.1007/s10571-007-9160-2> (2007).
- [43] Guo, H. T., Callaway, J. B. & Ting, J. P. Y. Inflammasomes: mechanism of action, role in disease, and therapeutics. *Nature Medicine* 21, 677-687, <https://doi.org/10.1038/nm.3893> (2015).
- [44] Lu, A. & Wu, H. Structural mechanisms of inflammasome assembly. *Febs Journal* 282, 435-444, <https://doi.org/10.1111/febs.13133> (2015).
- [45] Maelfait, J. et al. Stimulation of Toll-like receptor 3 and 4 induces interleukin-1? maturation by caspase-8. *The Journal of experimental medicine* 205, 1967-1973 (2008).
- [46] Stetson, D. B. & Medzhitov, R. Type I interferons in host defense. *Immunity* 25, 373-381 (2006).
- [47] Taub, D. D., Longo, D. L. & Murphy, W. J. Human interferon-inducible protein-10 induces mononuclear cell infiltration in mice and promotes the migration of human T lymphocytes into the peripheral tissues and human peripheral blood lymphocytes-SCID mice. *Blood* 87, 1423-1431 (1996).
- [48] Foy, B. D. et al. Probable non-vector-borne transmission of Zika virus, Colorado, USA. *Emerg Infect Dis* 17, 880-882, <https://doi.org/10.3201/eid1705.101939> (2011).
- [49] Haynes, M. Re: Thomas, L. C. et al. Pre-manipulative testing and the velocimeter. *Manual Therapy* (2007), 10:1016/j.math.2006.11.003. *Man Ther* 13, e4; author reply e5-6, doi:S1356-689X(07)00159-2 <https://doi.org/10.1016/j.math.2007.09.008> (2008).
- [50] La Morgia, C. et al. Association of the mtDNA m. 4171C > A/MT-ND1 mutation with both optic neuropathy and bilateral brainstem lesions. *BMC Neurol* 14, 116, <https://doi.org/10.1186/1471-2377-14-116> (2014).
- [51] Miao, E. A. et al. Cytoplasmic flagellin activates caspase-1 and secretion of interleukin 1beta via Ipaf. *Nat Immunol* 7, 569-575, <https://doi.org/10.1038/ni1344> (2006).
- [52] Doi, A. et al. Long RP' Tachycardia With Unusual Entrainment Responses: What Is the Mechanism? *J Cardiovasc Electrophysiol* 27, 1242-1244, <https://doi.org/10.1111/jce.12987> (2016).
- [53] Gabay, C., Lamacchia, C. & Palmer, G. IL-1 pathways in inflammation and human diseases. *Nat Rev Rheumatol* 6, 232-241, <https://doi.org/10.1038/nrrheum.2010.4> (2010).
- [54] Sivakumar, P. V. et al. Interleukin 18 is a primary mediator of the inflammation associated with dextran sulphate sodium induced colitis: blocking interleukin 18 attenuates intestinal damage. *Gut* 50, 812-820 (2002).
- [55] Suptawiwat, O. et al. A simple screening assay for receptor switching of avian influenza viruses. *Journal of Clinical Virology* 42, 186-189 (2008).
- [56] Levy, D. E. & Garc, A. The virus battles: IFN induction of the antiviral state and mechanisms of viral evasion. *Cytokine & growth factor reviews* 12, 143-156 (2001).
- [57] Schmid, S., Mordstein, M., Kochs, G. & García-Sastre, A. Transcription factor redundancy ensures induction of the antiviral state. *Journal of Biological Chemistry* 285, 42013-42022 (2010).
- [58] Haller, O. & Kochs, G. Human MxA protein: an interferon-induced dynamin-like GTPase with broad antiviral activity. *Journal of Interferon & Cytokine Research* 31, 79-87 (2011).
- [59] Yuan, J. et al. CXCL10 inhibits viral replication through recruitment of natural killer cells in coxsackievirus B3-induced myocarditis. *Circulation research* 104, 628-638 (2009).
- [60] Kumar, A. et al. Zika virus inhibits type?I interferon production and downstream signaling. *EMBO reports* 17, 1766-1775 (2016).