

# Terbium(III)-thiacalix[4]arene nanosensor for highly sensitive intracellular monitoring of temperature changes within the 303–313 K range

Zairov R.R., Dovzhenko A.P., Sapunova A.S., Voloshina A.D., Sarkanich K.A., Daminova A.G., Nizameev I.R., Lapaev D.V., Sudakova S.N., Podyachev S.N., Petrov K.A., Vomiero A., Mustafina A.R.

Kazan Federal University, 420008, Kremlevskaya 18, Kazan, Russia

---

## Abstract

© 2020, The Author(s). The work introduces hydrophilic PSS-[Tb<sub>2</sub>(TCAn)<sub>2</sub>] nanoparticles to be applied as highly sensitive intracellular temperature nanosensors. The nanoparticles are synthesized by solvent-induced nanoprecipitation of [Tb<sub>2</sub>(TCAn)<sub>2</sub>] complexes (TCAn - thiacalix[4]arenes bearing different upper-rim substituents: unsubstituted TCA1, tert-butyl-substituted TCA2, di- and tetra-brominated TCA3 and TCA4) with the use of polystyrenesulfonate (PSS) as stabilizer. The temperature responsive luminescence behavior of PSS-[Tb<sub>2</sub>(TCAn)<sub>2</sub>] within 293–333 K range in water is modulated by reversible changes derived from the back energy transfer from metal to ligand (M\* → T1) correlating with the energy gap between the triplet levels of ligands and resonant 5D<sub>4</sub> level of Tb<sup>3+</sup> ion. The lowering of the triplet level (T1) energies going from TCA1 and TCA2 to their brominated counterparts TCA3 and TCA4 facilitates the back energy transfer. The highest ever reported temperature sensitivity for intracellular temperature nanosensors is obtained for PSS-[Tb<sub>2</sub>(TCA4)<sub>2</sub>] (SI = 5.25% K<sup>-1</sup>), while PSS-[Tb<sub>2</sub>(TCA3)<sub>2</sub>] is characterized by a moderate one (SI = 2.96% K<sup>-1</sup>). The insignificant release of toxic Tb<sup>3+</sup> ions from PSS-[Tb<sub>2</sub>(TCAn)<sub>2</sub>] within heating/cooling cycle and the low cytotoxicity of the colloids point to their applicability in intracellular temperature monitoring. The cell internalization of PSS-[Tb<sub>2</sub>(TCAn)<sub>2</sub>] (n = 3, 4) marks the cell cytoplasm by green Tb<sup>3+</sup>-luminescence, which exhibits detectable quenching when the cell samples are heated from 303 to 313 K. The colloids hold unprecedented potential for in vivo intracellular monitoring of temperature changes induced by hyperthermia or pathological processes in narrow range of physiological temperatures.

<http://dx.doi.org/10.1038/s41598-020-77512-1>

---

## References

- [1] Bünzli, J. C. G. Lanthanide light for biology and medical diagnosis. *J. Lumin.* 170, 866–878 (2016). DOI: 10.1016/j.jlumin.2015.07.033
- [2] Wei, C. et al. Advances in luminescent lanthanide complexes and applications. *Sci. China Technol. Sci.* 61, 1265–1285 (2018). DOI: 10.1007/s11431-017-9212-7
- [3] Comby, S. et al. Lanthanide-functionalized nanoparticles as MRI and luminescent probes for sensing and/or imaging applications. *Inorg. Chem.* 53, 1867–1879 (2014). DOI: 10.1021/ic4023568

- [4] Zairov, R. R. et al. Dual red-NIR luminescent Eu Yb heterolanthanide nanoparticles as promising basis for cellular imaging and sensing. *Mater. Sci. Eng. C* 105, 110057 (2019). DOI: 10.1016/j.msec.2019.110057
- [5] Fedorenko, S. V. et al. Cellular imaging by green luminescence of Tb(III)-doped aminomodified silica nanoparticles. *Mater. Sci. Eng. C* 76, 551–558 (2017). DOI: 10.1016/j.msec.2017.03.106
- [6] Ferdinandus, et al. Facilely fabricated luminescent nanoparticle thermosensor for real-time microthermography in living animals. *ACS Sensors* 1, 1222–1227 (2016). DOI: 10.1021/acssensors.6b00320
- [7] Du, P., Luo, L. & Yu, J. S. Controlled synthesis and upconversion luminescence of Tm<sup>3+</sup>-doped NaYbF<sub>4</sub> nanoparticles for non-invasion optical thermometry. *J. Alloys Compd.* 739, 926–933 (2018). DOI: 10.1016/j.jallcom.2017.12.260
- [8] Du, P. & Yu, J. S. Synthesis of Er(III)/Yb(III)-doped BiF<sub>3</sub> upconversion nanoparticles for use in optical thermometry. *Microchim. Acta* 185, 237 (2018). DOI: 10.1007/s00604-018-2777-7
- [9] Savchuk, O. A. et al. Upconversion thermometry: a new tool to measure the thermal resistance of nanoparticles. *Nanoscale* 10, 6602–6610 (2018). DOI: 10.1039/C7NR08758F
- [10] Yao, L. et al. Regulation of morphologies and luminescence of β-NaGdF<sub>4</sub>: Yb<sup>3+</sup>, Er<sup>3+</sup> upconversion nanoparticles by hydrothermal method and their dual-mode thermometric properties. *Appl. Surf. Sci.* 466, 320–327 (2019). DOI: 10.1016/j.apsusc.2018.09.209
- [11] Kolesnikov, I. E. et al. Bifunctional heater-thermometer Nd<sup>3+</sup>-doped nanoparticles with multiple temperature sensing parameters. *Nanotechnology* 30, 145501 (2019). DOI: 10.1088/1361-6528/aafcb8
- [12] Zhang, Y. et al. Dually functioned core-shell NaYF<sub>4</sub>:Er<sup>3+</sup>/Yb<sup>3+</sup>@NaYF<sub>4</sub>:Tm<sup>3+</sup>/Yb<sup>3+</sup> nanoparticles as nanocalorifiers and nano-thermometers for advanced photothermal therapy. *Sci. Rep.* 7, 11849 (2017). DOI: 10.1038/s41598-017-11897-4
- [13] Wu, R. et al. α-NaYF<sub>4</sub>:Yb<sup>3+</sup>-Tm<sup>3+</sup>@CaF<sub>2</sub> nanocrystals for NIR-to-NIR temperature sensing. *Chem. Phys. Lett.* 667, 206–210 (2017). DOI: 10.1016/j.cplett.2016.10.040
- [14] Balabhadra, S., Debasu, M. L., Brites, C. D. S., Ferreira, R. A. S. & Carlos, L. D. Upconverting nanoparticles working as primary thermometers in different media. *J. Phys. Chem. C* 121, 13962–13968 (2017). DOI: 10.1021/acs.jpcc.7b04827
- [15] del Rosal, B., Ximendes, E., Rocha, U. & Jaque, D. In vivo luminescence nanothermometry: from materials to applications. *Adv. Opt. Mater.* 5, 1600508 (2017). DOI: 10.1002/adom.201600508
- [16] Pereira, A. F., Silva, J. F., Gouveia-Neto, A. S. & Jacinto, C. 1.319 Mm excited thulium doped nanoparticles for subtissue thermal sensing with deep penetration and high contrast imaging. *Sens. Actuators B Chem.* 238, 525–531 (2017). DOI: 10.1016/j.snb.2016.07.053
- [17] Skripka, A. et al. Double rare-earth nanothermometer in aqueous media: opening the third optical transparency window to temperature sensing. *Nanoscale* 9, 3079–3085 (2017). DOI: 10.1039/C6NR08472A
- [18] Fu, Y., Zhao, L., Guo, Y. & Yu, H. Up-conversion luminescence lifetime thermometry based on the 1G<sub>4</sub> state of Tm<sup>3+</sup> modulated by cross relaxation processes. *Dalt. Trans.* 48, 16034–16040 (2019). DOI: 10.1039/C9DT03452H
- [19] Kalinichev, A. A. et al. Near-infrared emitting YVO<sub>4</sub>:Nd<sup>3+</sup> nanoparticles for high sensitive fluorescence thermometry. *J. Lumin.* 195, 61–66 (2018). DOI: 10.1016/j.jlumin.2017.11.024
- [20] Yu, A., Zeng, Z., Luo, Q., Gao, J. & Wang, Q. Establishment of a new analytical platform for glucose detection based on a terbium containing silica hybrid nanosensor. *Appl. Surf. Sci.* 462, 883–889 (2018). DOI: 10.1016/j.apsusc.2018.08.162
- [21] Zhou, Z., Wang, Q., Zhang, C. C. & Gao, J. Molecular imaging of biothiols and: In vitro diagnostics based on an organic chromophore bearing a terbium hybrid probe. *Dalt. Trans.* 45, 7435–7442 (2016). DOI: 10.1039/C6DT00156D
- [22] Cao, J. et al. Optical thermometry based on up-conversion luminescence behavior of self-crystallized K<sub>3</sub>YF<sub>6</sub>:Er<sup>3+</sup> glass ceramics. *Sens. Actuators B Chem.* 224, 507–513 (2016). DOI: 10.1016/j.snb.2015.10.087
- [23] Rocha, U. et al. Neodymium-doped LaF<sub>3</sub> nanoparticles for fluorescence bioimaging in the second biological window. *Small* 10, 1141–1154 (2014). DOI: 10.1002/smll.201301716
- [24] Savchuk, O. A. et al. Ho, Yb:KLu(WO<sub>4</sub>)<sub>2</sub> nanoparticles: a versatile material for multiple thermal sensing purposes by luminescent thermometry. *J. Phys. Chem. C* 119, 18546–18558 (2015). DOI: 10.1021/acs.jpcc.5b03766
- [25] Pudovkin, M. S. et al. The comparison of Pr<sup>3+</sup>:LaF<sub>3</sub> and Pr<sup>3+</sup>:LiYF<sub>4</sub> luminescent nano- and microthermometer performances. *J. Nanoparticle Res.* 21, 266 (2019). DOI: 10.1007/s11051-019-4713-0
- [26] Yao, L. et al. Simultaneous enhancement of upconversion luminescence and thermometric property of upconversion nanoparticles by tuning crystal field. *J. Lumin.* 211, 144–149 (2019). DOI: 10.1016/j.jlumin.2019.03.030
- [27] Runowski, M., Woźny, P., Martín, I. R., Lavín, V. & Lis, S. Praseodymium doped YF<sub>3</sub>:Pr<sup>3+</sup> nanoparticles as optical thermometer based on luminescence intensity ratio (LIR)—studies in visible and NIR range. *J. Lumin.* 214, 116571 (2019). DOI: 10.1016/j.jlumin.2019.116571

- [28] Zhou, K. et al. Fabrication and optical thermometry of transparent glass-ceramics containing Ag@NaGdF<sub>4</sub>:Er<sup>3+</sup> core-shell nanocrystals. *J. Am. Ceram. Soc.* 102, 6564–6574 (2019). DOI: 10.1111/jace.16519
- [29] Brites, C. D. S. et al. A luminescent molecular thermometer for long-term absolute temperature measurements at the nanoscale. *Adv. Mater.* 22, 4499–4504 (2010). DOI: 10.1002/adma.201001780
- [30] Brites, C. D. S. et al. Lanthanide-based luminescent molecular thermometers. *New J. Chem.* 35, 1177–1183 (2011). DOI: 10.1039/c0nj01010c
- [31] Wang, X. D., Wolfbeis, O. S. & Meier, R. J. Luminescent probes and sensors for temperature. *Chem. Soc. Rev.* 42, 7834–7869 (2013). DOI: 10.1039/c3cs60102a
- [32] Brites, C. D. S., Lima, P. P. & Carlos, L. D. Tuning the sensitivity of Ln<sup>3+</sup>-based luminescent molecular thermometers through ligand design. *J. Lumin.* 169, 497–502 (2016). DOI: 10.1016/j.jlumin.2015.01.025
- [33] Peng, H. et al. Luminescent europium(III) nanoparticles for sensing and imaging of temperature in the physiological range. *Adv. Mater.* 22, 716–719 (2010). DOI: 10.1002/adma.200901614
- [34] Brites, C. D. S. et al. Thermometry at the nanoscale. *Nanoscale* 4, 4799–4829 (2012). DOI: 10.1039/c2nr30663h
- [35] Alberto, G., Caputo, G., Viscardi, G., Coluccia, S. & Martra, G. Molecular engineering of hybrid dye-silica fluorescent nanoparticles: Influence of the dye structure on the distribution of fluorophores and consequent photoemission brightness. *Chem. Mater.* 24, 2792–2801 (2012). DOI: 10.1021/cm301308g
- [36] Gubala, V., Giovannini, G., Kunc, F., Monopoli, M. P. & Moore, C. J. Dye-Doped Silica Nanoparticles: Synthesis, Surface Chemistry and Bioapplications. *Cancer Nanotechnology* vol. 11 (Springer Vienna, 2020).
- [37] Zairov, R. R. et al. Polyelectrolyte-coated ultra-small nanoparticles with Tb(III)-centered luminescence as cell labels with unusual charge effect on their cell internalization. *Mater. Sci. Eng. C* 95, 166–173 (2019). DOI: 10.1016/j.msec.2018.10.084
- [38] Podyachev, S. N. et al. Structural and photophysical properties of Tb<sup>3+</sup>-tetra-1,3-diketonate complexes controlled by calix[4]arene-tetrathiacalix[4]arene scaffolds. *Dalt. Trans.* 48, 3930–3940 (2019). DOI: 10.1039/C9DT00286C
- [39] Podyachev, S. S. S. et al. A simple synthetic approach to enhance the thermal luminescence sensitivity of Tb<sup>3+</sup> complexes with thiacalix[4]arene derivatives through upper-rim bromination. *Dalt. Trans.* 49, 8293–8313. DOI: 10.1039/d0dt00709a (2020).
- [40] Higuchi, Y. Fluorescent chemo-sensor for metal cations based on Thiacalix[4]arenes modified with dansyl moieties at the lower rim. *Tetrahedron* 56, 4659–4666 (2000). DOI: 10.1016/S0040-4020(00)00377-X
- [41] Kumagai, H. et al. Facile synthesis of p-tert-butylthiacalix[4]arene by the reaction of p-tert-butylphenol with elemental sulfur in the presence of a base. *Tetrahedron Lett.* 38, 3971–3972 (1997). DOI: 10.1016/S0040-4039(97)00792-2
- [42] Desroches, C., Lopes, C., Kessler, V. & Parola, S. Design and synthesis of multifunctional thiacalixarenes and related metal derivatives for the preparation of sol-gel hybrid materials with non-linear optical properties. *Dalt. Trans.* 10.1039/B210252H (2003). DOI: 10.1039/B210252H
- [43] Zairov, R. et al. Paramagnetic relaxation enhancement in hydrophilic colloids based on Gd(III) complexes with tetrathia- and Calix[4]arenes. *J. Phys. Chem. C* 124, 4320–4329 (2020). DOI: 10.1021/acs.jpcc.0c00312
- [44] Zairov, R. et al. Hydration number: crucial role in nuclear magnetic relaxivity of Gd(III) chelate-based nanoparticles. *Sci. Rep.* 7, 1–10 (2017). DOI: 10.1038/s41598-016-0028-x
- [45] Richardson, F. S. Terbium(III) and europium(III) ions as luminescent probes and stains for biomolecular systems. *Chem. Rev.* 82, 541–552 (1982). DOI: 10.1021/cr00051a004
- [46] Horrocks, W., Deuk, W. & Sudnick, D. R. Lanthanide ion probes of structure in biology laser-induced luminescence decay constants provide a direct measure of the number of metal-coordinated water molecules. *J. Am. Chem. Soc.* 101, 334–340 (1979). DOI: 10.1021/ja00496a010
- [47] Latva, M. et al. Correlation between the lowest triplet state energy level of the ligand and lanthanide(III) luminescence quantum yield. *J. Lumin.* 75, 149–169 (1997). DOI: 10.1016/S0022-2313(97)00113-0
- [48] Gusev, A. N. et al. Synthesis, structure and luminescence studies of Eu(III), Tb(III), Sm(III), Dy(III) cationic complexes with acetylacetonate and bis(5-(pyridine-2-yl)-1, 2, 4-triazol-3-yl)propane. *Inorganica Chim. Acta* 406, 279–284 (2013). DOI: 10.1016/j.ica.2013.04.006
- [49] Ma, H. Y. et al. Roles of reactive oxygen species (ROS) in the photocatalytic degradation of pentachlorophenol and its main toxic intermediates by TiO<sub>2</sub>/UV. *J. Hazard. Mater.* 369, 719–726 (2019). DOI: 10.1016/j.jhazmat.2019.02.080
- [50] Steinmetz, Z. et al. Biodegradation and photooxidation of phenolic compounds in soil—a compound-specific stable isotope approach. *Chemosphere* 230, 210–218 (2019). DOI: 10.1016/j.chemosphere.2019.05.030
- [51] Shamsutdinova, N. et al. Tuning magnetic relaxation properties of “hard cores” in core-shell colloids by modification of “soft shell”. *Colloids Surf. B Biointerfaces* 162, 52–59 (2018). DOI: 10.1016/j.colsurfb.2017.10.070

- [52] Liu, N., Tang, M. & Ding, J. The interaction between nanoparticles-protein corona complex and cells and its toxic effect on cells. *Chemosphere* 245, 125624 (2020). DOI: 10.1016/j.chemosphere.2019.125624
- [53] Shahabi, S., Treccani, L., Dringen, R. & Rezwan, K. Modulation of silica nanoparticle uptake into human osteoblast cells by variation of the ratio of amino and sulfonate surface groups: effects of serum. *ACS Appl. Mater. Interfaces* 7, 13821–13833 (2015). DOI: 10.1021/acsami.5b01900