

Δccr5 Genotype Is Associated with Mild Form of Nephropathia Epidemica

Kletenkov K., Martynova E., Davidyuk Y., Kabwe E., Shamsutdinov A., Garanina E., Shakirova V., Khaertynova I., Anokhin V., Tarlinton R., Rizvanov A., Khaiboullina S., Morzunov S.
Kazan Federal University, 420008, Kremlevskaya 18, Kazan, Russia

Abstract

© 2019 by the authors. Licensee MDPI, Basel, Switzerland. Nephropathia Epidemica (NE), a mild form of hemorrhagic fever with renal syndrome (HFRS) and linked to hantavirus infection, is endemic in the Republic of Tatarstan. Several genetic markers of HFRS severity have been identified previously, including human leukocyte antigen (HLA) complexes and nucleotide polymorphism in the tumor necrosis factor alpha (TNF α) gene. Still, our understanding of the genetic markers of NE severity remains incomplete. The frequency of the C-C chemokine receptor type 5 (CCR5) gene wild type and gene with 32-base-pair deletion (Δ 32CCR5) genotypes in 98 NE samples and 592 controls was analyzed using PCR. Along with the serum levels of 94 analytes, a lack of differences in the CCR5 genotype distribution between NE cases and the general population suggests that the CCR5 genotype does not affect susceptibility to hantavirus infection. However, in NE cases, significant variation in the serum levels of the host matrix metalloproteases between functional CCR5 homozygous and Δ 32CCR5 heterozygous patients was detected. Also, the oliguric phase was longer, while thrombocyte counts were lower in functional CCR5 homozygous as compared to heterozygous NE cases. Our data, for the first time, presents the potential role of the CCR5 receptor genotype in NE pathogenesis. Our data suggests that NE pathogenesis in functional CCR5 homozygous and heterozygous NE patients differs, where homozygous cases may have more disintegration of the extracellular matrix and potentially more severe disease.

<http://dx.doi.org/10.3390/v11070675>

Keywords

CCR5, Cytokines, Hemorrhagic fever with renal syndrome, Matrix metalloprotease, Nephropathia epidemica, Δ 32CCR5

References

- [1] Khismatullina, N.A.; Karimov, M.M.; Khaertynov, K.S.; Shuralev, E.A.; Morzunov, S.P.; Khaertynova, I.M.; Ivanov, A.A.; Milova, I.V.; Khakimzyanova, M.B.; Sayfullina, G.; et al. Epidemiological dynamics of nephropathia epidemica in the Republic of Tatarstan, Russia, during the period of 1997–2013. *Epidemiol. Infect.* 2016, 144, 618–626. [CrossRef] [PubMed]
- [2] Rasche, F.M.; Uhel, B.; Kruger, D.H.; Karges, W.; Czock, D.; Hampl, W.; Keller, F.; Meisel, H.; von Muller, L. Thrombocytopenia and acute renal failure in Puumala hantavirus infections. *Emerg. Infect. Dis.* 2004, 10, 1420–1425. [CrossRef] [PubMed]

- [3] Temonen, M.; Mustonen, J.; Helin, H.; Pasternack, A.; Vaheri, A.; Holthofer, H. Cytokines, adhesion molecules, and cellular infiltration in nephropathia epidemica kidneys: An immunohistochemical study. *Clin. Immunol. Immunopathol.* 1996, 78, 47-55. [CrossRef] [PubMed]
- [4] Sokol, C.L.; Luster, A.D. The chemokine system in innate immunity. *Cold Spring Harb. Perspect. Biol.* 2015, 7. [CrossRef] [PubMed]
- [5] Martynova, E.V.; Valiullina, A.H.; Gusev, O.A.; Davidiyuk, Y.N.; Garanina, E.E.; Shakirova, V.G.; Khaertynova, I.; Anokhin, V.A.; Rizvanov, A.A.; Khaiboullina, S.F. High Triglycerides Are Associated with Low Thrombocyte Counts and High VEGF in Nephropathia Epidemica. *J. Immunol. Res.* 2016, 2016, 8528270. [CrossRef] [PubMed]
- [6] Wu, L.; LaRosa, G.; Kassam, N.; Gordon, C.J.; Heath, H.; Ruffing, N.; Chen, H.; Humblias, J.; Samson, M.; Parmentier, M.; et al. Interaction of chemokine receptor CCR5 with its ligands: Multiple domains for HIV-1 gp120 binding and a single domain for chemokine binding. *J. Exp. Med.* 1997, 186, 1373-1381. [CrossRef] [PubMed]
- [7] Schall, T.J.; Bacon, K.; Toy, K.J.; Goeddel, D.V. Selective attraction of monocytes and T lymphocytes of the memory phenotype by cytokine RANTES. *Nature* 1990, 347, 669-671. [CrossRef]
- [8] Glass, W.G.; Lim, J.K.; Cholera, R.; Pletnev, A.G.; Gao, J.L.; Murphy, P.M. Chemokine receptor CCR5 promotes leukocyte trafficking to the brain and survival in West Nile virus infection. *J. Exp. Med.* 2005, 202, 1087-1098. [CrossRef]
- [9] Hull, J.; Rowlands, K.; Lockhart, E.; Moore, C.; Sharland, M.; Kwiatkowski, D. Variants of the chemokine receptor CCR5 are associated with severe bronchiolitis caused by respiratory syncytial virus. *J. Infect. Dis.* 2003, 188, 904-907. [CrossRef] [PubMed]
- [10] Mori, M.; Rothman, A.L.; Kurane, I.; Montoya, J.M.; Nolte, K.B.; Norman, J.E.; Waite, D.C.; Koster, F.T.; Ennis, F.A. High levels of cytokine-producing cells in the lung tissues of patients with fatal hantavirus pulmonary syndrome. *J. Infect. Dis.* 1999, 179, 295-302. [CrossRef] [PubMed]
- [11] Sironen, T.; Klingstrom, J.; Vaheri, A.; Andersson, L.C.; Lundkvist, A.; Plyusnin, A. Pathology of Puumala hantavirus infection in macaques. *PLoS ONE* 2008, 3, e3035. [CrossRef] [PubMed]
- [12] Zaki, S.R.; Greer, P.W.; Coffield, L.M.; Goldsmith, C.S.; Nolte, K.B.; Foucar, K.; Feddersen, R.M.; Zumwalt, R.E.; Miller, G.L.; Khan, A.S.; et al. Hantavirus pulmonary syndrome. Pathogenesis of an emerging infectious disease. *Am. J. Pathol.* 1995, 146, 552-579. [PubMed]
- [13] Rugeles, M.T.; Solano, F.; Diaz, F.J.; Bedoya, V.I.; Patino, P.J. Molecular characterization of the CCR 5 gene in seronegative individuals exposed to human immunodeficiency virus (HIV). *J. Clin. Virol.* 2002, 23, 161-169. [CrossRef]
- [14] R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing. Available online: <https://www.R-project.org/>(accessed on 1 April 2017).
- [15] RStudio: Integrated Development for R; RStudio, Inc.: Boston, MA, USA, 2015.
- [16] The "Comprehensive R Archive Network" (CRAN). Available online: <https://CRAN.R-project.org/>(accessed on 23 July 2019).
- [17] Wickham, H. *ggplot2: Elegant Graphics for Data Analysis*; Springer: New York, NY, USA, 2009.
- [18] Dick, J.M. Chemical composition and the potential for proteomic transformation in cancer, hypoxia, and hyperosmotic stress. *PeerJ* 2017, 5, e3421. [CrossRef] [PubMed]
- [19] Kuhn, M.; Jed Wing, S.W.; Williams, A.; Keefer, C.; Engelhardt, A.; Cooper, T.; Mayer, Z.; Kenkel, B.; Benesty, M.; the R Core Team; et al. Misc Functions for Training and Plotting Classification and Regression Models. The "Comprehensive R Archive Network" (CRAN). Available online: <https://CRAN.R-project.org/>(accessed on 22 July 2019).
- [20] caret: Classification and Regression Training. R Package Version 6.0-78. Available online: <https://CRAN.R-project.org/package=caret> (accessed on 1 April 2017).
- [21] Wickham, H. Reshaping Data with the reshape Package. *J. Stat. Softw.* 2017, 21, 1-20.
- [22] Outinen, T.K.; Laine, O.K.; Makela, S.; Porsti, I.; Huhtala, H.; Vaheri, A.; Mustonen, J. Thrombocytopenia associates with the severity of inflammation and variables reflecting capillary leakage in Puumala Hantavirus infection, an analysis of 546 Finnish patients. *Infect. Dis. (Lond.)* 2016, 48, 682-687. [CrossRef] [PubMed]
- [23] Kristiansen, M.; Graversen, J.H.; Jacobsen, C.; Sonne, O.; Hoffman, H.J.; Law, S.K.; Moestrup, S.K. Identification of the haemoglobin scavenger receptor. *Nature* 2001, 409, 198-201. [CrossRef]
- [24] Doni, A.; Peri, G.; Chieppa, M.; Allavena, P.; Pasqualini, F.; Vago, L.; Romani, L.; Garlanda, C.; Mantovani, A. Production of the soluble pattern recognition receptor PTX3 by myeloid, but not plasmacytoid, dendritic cells. *Eur. J. Immunol.* 2003, 33, 2886-2893. [CrossRef]
- [25] Marsac, D.; García, S.; Fournet, A.; Aguirre, A.; Pino, K.; Ferres, M.; Kalergis, A.M.; Lopez-Lastra, M.; Veas, F. Infection of human monocyte-derived dendritic cells by ANDES Hantavirus enhances pro-inflammatory state, the secretion of active MMP-9 and indirectly enhances endothelial permeability. *Virol. J.* 2011, 8, 223. [CrossRef]

- [26] Easterbrook, J.D.; Klein, S.L. Corticosteroids modulate Seoul virus infection, regulatory T cell responses, and MMP-9 expression in male, but not female, Norway rats. *J. Gen. Virol.* 2008, 89, 2723. [CrossRef]
- [27] Khaiboullina, S.F.; Levis, S.; Morzunov, S.P.; Martynova, E.V.; Anokhin, V.A.; Gusev, O.A.; St Jeor, S.C.; Lombardi, V.C.; Rizvanov, A.A. Serum cytokine profiles differentiating hemorrhagic fever with renal syndrome and hantavirus pulmonary syndrome. *Front. Immunol.* 2017, 8, 567. [CrossRef] [PubMed]
- [28] Wang, M.; Wang, J.; Wang, T.; Li, J.; Hui, L.; Ha, X. Thrombocytopenia as a predictor of severe acute kidney injury in patients with Hantaan virus infections. *PLoS ONE* 2013, 8, e53236. [CrossRef] [PubMed]
- [29] Pothapregada, S.; Kamalakannan, B.; Thulasingham, M. Role of platelet transfusion in children with bleeding in dengue fever. *J. Vector Borne Dis.* 2015, 52, 304–308. [PubMed]
- [30] Perrier, P.; Martinez, F.O.; Locati, M.; Bianchi, G.; Nebuloni, M.; Vago, G.; Bazzoni, F.; Sozzani, S.; Allavena, P.; Mantovani, A. Distinct transcriptional programs activated by interleukin-10 with or without lipopolysaccharide in dendritic cells: Induction of the B cell-activating chemokine, CXC chemokine ligand 13. *J. Immunol.* 2004, 172, 7031–7042. [CrossRef] [PubMed]
- [31] Viola, A.; Luster, A.D. Chemokines and their receptors: Drug targets in immunity and inflammation. *Annu. Rev. Pharmacol. Toxicol.* 2008, 48, 171–197. [CrossRef] [PubMed]
- [32] Kohlmeier, J.E.; Miller, S.C.; Smith, J.; Lu, B.; Gerard, C.; Cookenham, T.; Roberts, A.D.; Woodland, D.L. The chemokine receptor CCR5 plays a key role in the early memory CD8+ T cell response to respiratory virus infections. *Immunity* 2008, 29, 101–113. [CrossRef]
- [33] Page-McCaw, A.; Ewald, A.J.; Werb, Z. Matrix metalloproteinases and the regulation of tissue remodelling. *Nat. Rev. Mol. Cell. Biol.* 2007, 8, 221–233. [CrossRef] [PubMed]
- [34] Whitelock, J.M.; Murdoch, A.D.; Iozzo, R.V.; Underwood, P.A. The degradation of human endothelial cell-derived perlecan and release of bound basic fibroblast growth factor by stromelysin, collagenase, plasmin, and heparanases. *J. Biol. Chem.* 1996, 271, 10079–10086. [CrossRef]
- [35] Abraham, M.; Shapiro, S.; Karni, A.; Weiner, H.L.; Miller, A. Gelatinases (MMP-2 and MMP-9) are preferentially expressed by Th1 vs. Th2 cells. *J. Neuroimmunol.* 2005, 163, 157–164. [CrossRef]
- [36] Bini, A.; Itoh, Y.; Kudryk, B.J.; Nagase, H. Degradation of cross-linked fibrin by matrix metalloproteinase 3 (stromelysin 1): Hydrolysis of the gamma Gly 404-Ala 405 peptide bond. *Biochemistry* 1996, 35, 13056–13063. [CrossRef]
- [37] Krampert, M.; Bloch, W.; Sasaki, T.; Bugnon, P.; Rulicke, T.; Wolf, E.; Aumailley, M.; Parks, W.C.; Werner, S. Activities of the matrix metalloproteinase stromelysin-2 (MMP-10) in matrix degradation and keratinocyte organization in wounded skin. *Mol. Biol. Cell.* 2004, 15, 5242–5254. [CrossRef] [PubMed]
- [38] Jovanovic, D.V.; di Battista, J.A.; Martel-Pelletier, J.; Reboul, P.; He, Y.; Jolicœur, F.C.; Pelletier, J.P. Modulation of TIMP-1 synthesis by antiinflammatory cytokines and prostaglandin E2 in interleukin 17 stimulated human monocytes/macrophages. *J. Rheumatol.* 2001, 28, 712–718. [PubMed]
- [39] Van Hamburg, J.P.; Asmawidjaja, P.S.; Davelaar, N.; Mus, A.M.; Colin, E.M.; Hazes, J.M.; Dolhain, R.J.; Lubberts, E. Th17 cells, but not Th1 cells, from patients with early rheumatoid arthritis are potent inducers of matrix metalloproteinases and proinflammatory cytokines upon synovial fibroblast interaction, including autocrine interleukin-17A production. *Arthritis Rheum.* 2011, 63, 73–83. [CrossRef] [PubMed]
- [40] Homey, B.; Alenius, H.; Muller, A.; Soto, H.; Bowman, E.P.; Yuan, W.; McEvoy, L.; Lauerma, A.I.; Assmann, T.; Bunemann, E.; et al. CCL27-CCR10 interactions regulate T cell-mediated skin inflammation. *Nat. Med.* 2002, 8, 157–165. [CrossRef] [PubMed]